

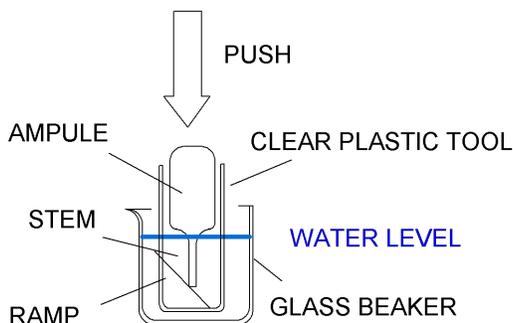
**PaSEC**  
**Colorimeter**  
**Instructions**  
**DR850**



## Dissolved Oxygen Test – Colorimeter – Instruction #4 – Replacement: Using the AccuVac Snapper Tool

### Breaking the Dissolved Oxygen Ampule Tip:

Use the glass beaker and the AccuVac Snapper (clear plastic tool) to break the stem on the ampule. Illustration #4, p. 453 in the PaSEC Colorimeter DR850 Instructions is misleading. The schematic drawing below illustrates the process.

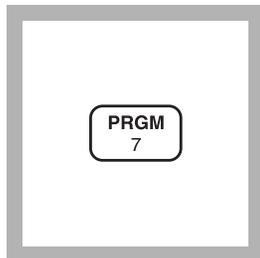


### Instruction #4, p.453 Replacement:

- Fill the glass beaker with about 40 ml. of sample water. (Instruction #3)
- Grip the open end of the Snapper tool between index and middle fingers with open end up.
- Gently slip the ampule into the Snapper tool, point first, until tip **touches** the ramp.
- Lower the ribbed end of Snapper tool into sample, until the ampule shoulder is wet.
- Press on the flat end of ampule with thumb until the tip snaps, and allow the ampule to fill. The vacuum in the ampule will draw sample water into and fill the ampule. As the reagent reacts with dissolved oxygen in the sample, a color change from yellow to purple will occur if oxygen is present.
- Remove ampule from sample in glass beaker **without inverting**, and cap, as directed in Instruction #5, p.453.
- Continue with instruction #5, p.453, in the PaSEC Colorimeter DR850 Instructions for the Dissolved Oxygen Test.
- The ampule is to be picked up and wiped free of fingerprints and water drops with a soft cloth before inserting into colorimeter for Dissolved Oxygen reading determination.
- Knock out ampule tip from Snapper tool into suitable waste container.
- Rinse glass beaker and Snapper tool with deionized water before storing.

**OXYGEN, DISSOLVED, High Range (0 to 15.0 mg/L O<sub>2</sub>)****HRDO Method**

For water and wastewater

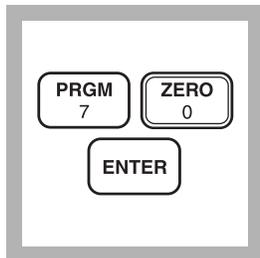


1. Enter the stored program number for dissolved oxygen, high range.

Press: **PRGM**

The display will show:

**PRGM ?**

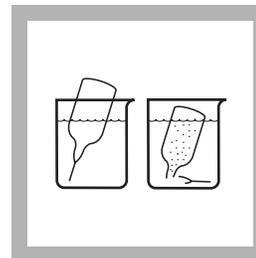


2. Press: **70 ENTER**

The display will show **mg/L, O<sub>2</sub>** and the **ZERO** icon.

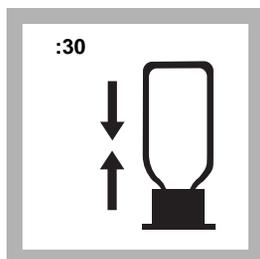


3. Fill a sample cell (the blank) with at least 10 mL of sample. Fill a blue ampul cap with sample. Collect at least 40 mL of sample in a 50-mL beaker.



4. Fill a High Range Dissolved Oxygen AccuVac Ampul with sample.

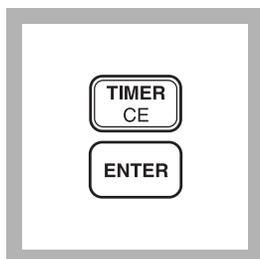
*Note: Keep the tip immersed while the ampul fills completely.*



5. Without inverting the ampul, immediately place the ampul cap that has been filled with sample securely over the tip of the ampul. Shake for about 30 seconds.

*Note: Accuracy is not affected by undissolved powder.*

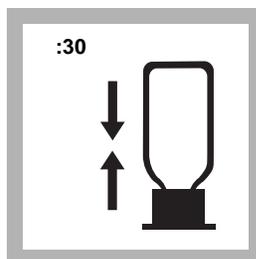
*Note: The cap prevents contamination with atmospheric oxygen.*



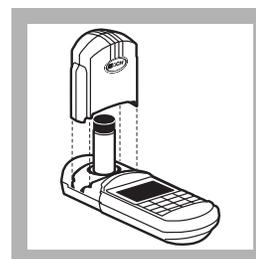
6. Press: **TIMER ENTER**

A 2-minute reaction period will begin.

*Note: The two-minute period allows oxygen which was degassed during aspiration to redissolve in the sample and react.*



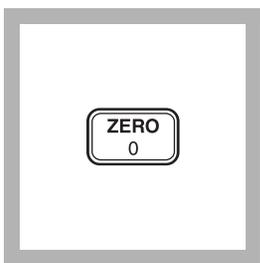
7. When the timer beeps, shake the ampul for 30 seconds.



8. Place the blank into the cell holder. Tightly cover the sample cell with the instrument cap.

## OXYGEN, DISSOLVED, High Range, continued

---

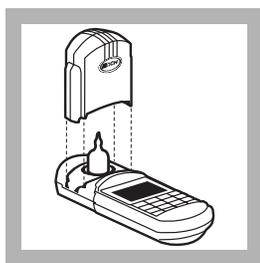


**9. Press: ZERO**

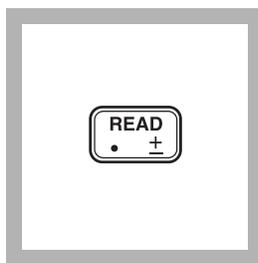
The cursor will move to the right, then the display will show:

**0.0 mg/L O<sub>2</sub>**

*Note: If Reagent Blank Correction is on, the display may flash "limit". See Section 1.*



**10. Place the AccuVac ampul into the cell holder. Tightly cover the ampul with the instrument cap. Wait approximately 30 seconds for the air bubbles to disperse from the light path.**



**11. Press: READ**

The cursor will move to the right, then the result in mg/L O<sub>2</sub> will be displayed.

*Note: Standard Adjust may be performed using a prepared standard (see Section 1).*

---

### Sampling and Storage

The main consideration in sampling with the High Range Dissolved Oxygen AccuVac Ampul is to prevent the sample from becoming contaminated with atmospheric oxygen. This is accomplished by capping the ampul with an ampul cap in the interval between breaking open the ampul and reading the absorbance. If the ampul is securely capped, it should be safe from contamination for several hours. The absorbance will decrease by approximately 3% during the first hour and will not change significantly afterwards.

Sampling and sample handling are important considerations in obtaining meaningful results. The dissolved oxygen content of the water being tested can be expected to change with depth, turbulence, temperature, sludge deposits, light, microbial action, mixing, travel time and other factors. A single dissolved oxygen test rarely reflects the accurate over-all condition of a body of water. Several samples taken at different times, locations and depths are recommended for most reliable results. Samples must be tested immediately upon collection although only a small error results if the absorbance reading is taken several hours later.

## OXYGEN, DISSOLVED, High Range, continued

---

### Accuracy Check

The results of this procedure may be compared with the results of a dissolved oxygen meter (Cat. No. 51815-01).

### Method Performance

#### Precision

In a single laboratory, using a standard solution of 8.0 mg/L O<sub>2</sub> determined by the Winkler method and two representative lots of reagent with the instrument, a single operator obtained a standard deviation of ±0.41 mg/L O<sub>2</sub>.

#### Estimated Detection Limit

The estimated detection limit for program 70 is 0.10 mg/L O<sub>2</sub>. For more information on the estimated detection limit, see *Section 1*.

### Interferences

Interfering Substance	Interference Levels and Treatments
Cr <sup>3+</sup>	Greater than 10 mg/L
Cu <sup>2+</sup>	Greater than 10 mg/L
Fe <sup>2+</sup>	Greater than 10 mg/L
Mg <sup>2+</sup>	Magnesium is commonly present in seawater and causes a negative interference. If the sample contains more than 50% seawater, the oxygen concentration obtained by this method will be 25% less than the true oxygen concentration. If the sample contains less than 50% seawater, the interference will be less than 5%.
Mn <sup>2+</sup>	Greater than 10 mg/L
Ni <sup>2+</sup>	Greater than 10 mg/L
NO <sub>2</sub> <sup>-</sup>	Greater than 10 mg/L

### Summary of Method

The High Range Dissolved Oxygen AccuVac Ampul contains reagent vacuum sealed in a 12-mL ampul. When the AccuVac ampul is broken open in a sample containing dissolved oxygen, a yellow color forms, which turns purple as the oxygen reacts with the reagent. The color developed is proportional to the concentration of dissolved oxygen.

## OXYGEN, DISSOLVED, High Range, continued

---

### REQUIRED REAGENTS

Description	Quantity Required Per Test	Unit	Cat. No.
High Range Dissolved Oxygen AccuVac Ampuls, with 2 reusable ampul caps .....	1 ampul .....	25/pkg.....	25150-25

### REQUIRED APPARATUS

Beaker, 50 mL.....	1 .....	each.....	500-41H
Caps, ampul, blue.....	varies .....	25/pkg.....	1731-25
Sample Cell, 10-20-25 mL, w/ cap.....	1 .....	6/pkg.....	24019-06

### OPTIONAL REAGENTS AND APPARATUS

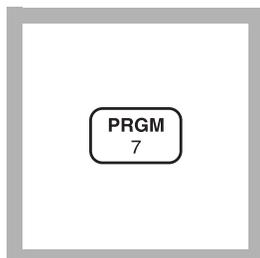
AccuVac Dissolved Oxygen Sampler .....		each.....	24051-00
AccuVac Snapper Kit.....		each.....	24052-00
AccuVac Drainer.....		each.....	41036-00
BOD bottle and stopper, 300 mL.....		each.....	621-00
Dissolved Oxygen Meter, Portable HQ 10 .....		each.....	51815-01
Dissolved Oxygen Reagent Set (Buret Method).....	100 tests.....		23514-00
Dissolved Oxygen Reagent Set (Digital Titrator Method) .....	50 tests.....		22722-00

Dissolved oxygen may also be determined by titrimetric methods.  
Request Publication 8042 for additional information.

### *For Technical Assistance, Price and Ordering*

In the U.S.A.—Call 800-227-4224

Outside the U.S.A.—Contact the Hach office or distributor serving you.

**NITRATE, High Range (0 to 30.0 mg/L NO<sub>3</sub><sup>-</sup>-N) For water, wastewater, and seawater\*****Cadmium Reduction Method (Using Powder Pillows or AccuVac Ampuls)  
Using Powder Pillows**

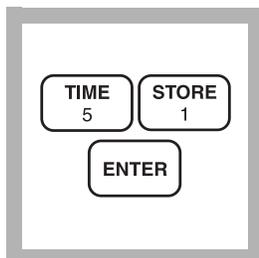
1. Enter the stored program number for high range nitrate nitrogen (NO<sub>3</sub><sup>-</sup>-N) powder pillows.

Press: **PRGM**

The display will show:

**PRGM ?**

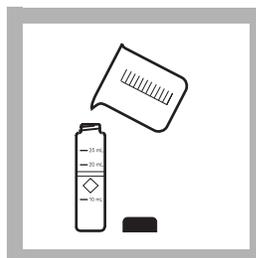
*Note: For most accurate results, perform a Reagent Blank Correction using deionized water (see Section 1).*



2. Press: **51 ENTER**

The display will show **mg/L, NO<sub>3</sub>-N** and the **ZERO** icon.

*Note: For alternate forms (NO<sub>3</sub>), press the **CONC** key.*



3. Fill a sample cell with 10 mL of sample.

*Note: Adjust the pH of stored samples before analysis.*

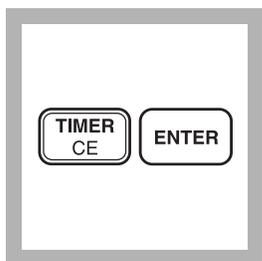


4. Add the contents of one NitraVer 5 Nitrate Reagent Powder Pillow to the sample cell (the prepared sample). Cap the sample cell.

*Note: It is important to remove all of the powder from the foil pillow. Tap the pillow until no more powder pours out.*

\* Seawater requires a manual calibration; see Interferences.

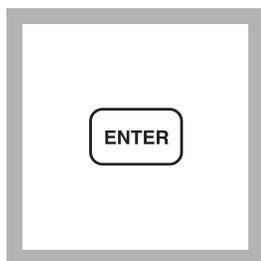
## NITRATE, High Range, continued



**5. Press:**  
**TIMER ENTER**

A one-minute reaction period will begin. Shake the sample cell vigorously until the timer beeps.

*Note: It is important to shake the cell vigorously. Shaking time and technique influence color development. For most accurate results, do successive tests on a standard solution and adjust the shaking time to obtain the correct result.*



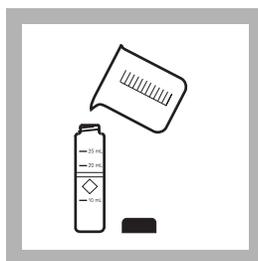
**6. After the timer beeps, the display will show:**  
**5:00 TIMER 2**

Press: **ENTER**

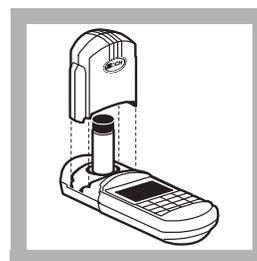
A five-minute reaction period will begin.

*Note: A deposit will remain after the reagent dissolves and will not affect test results.*

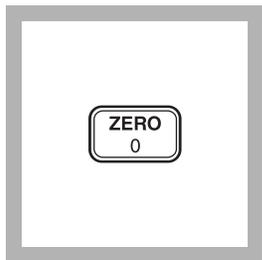
*Note: An amber color will develop if nitrate nitrogen is present.*



**7. Fill another cell with 10 mL of sample (the blank). Wipe off any fingerprints or liquid.**



**8. Place the blank into the cell holder. Tightly cover the sample cell with the instrument cap.**

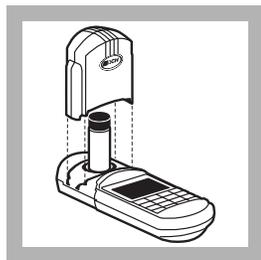


**9. When the timer beeps, press ZERO.**

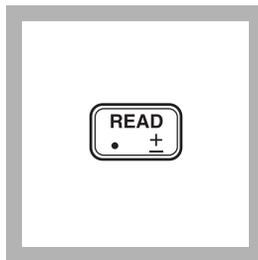
The cursor will move to the right, then the display will show:

**0.0 mg/L NO<sub>3</sub>-N**

*Note: If Reagent Blank Correction is on, the display may flash "limit". See Section 1.*



**10. Place the prepared sample into the cell holder. Tightly cover the sample cell with the instrument cap.**



**11. Press: READ**

The cursor will move to the right, then the result in mg/L NO<sub>3</sub>-N (or alternate form) will be displayed.

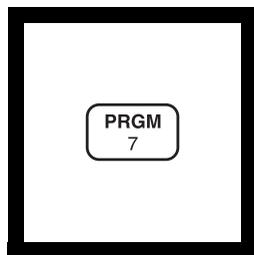
*Note: Use of the Standard Adjust feature for each new lot of reagent is highly recommended. See Accuracy Check.*

*Note: Rinse the sample cell immediately after use to remove all cadmium particles. Save the spent sample for proper hazardous waste disposal for cadmium.*

# NITRATE, Low Range (0 to 0.50 mg/L NO<sub>3</sub><sup>-</sup>-N)

For water, wastewater and seawater\*

## Cadmium Reduction Method



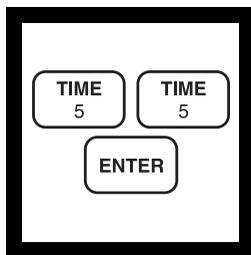
**1.** Enter the stored program number for low range nitrate nitrogen (NO<sub>3</sub><sup>-</sup>-N).

Press: **PRGM**

The display will show:

**PRGM ?**

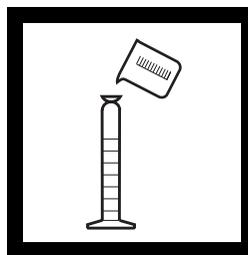
*Note:* For most accurate results, perform a Reagent Blank Correction using deionized water (see Section 1).



**2.** Press: **55 ENTER**

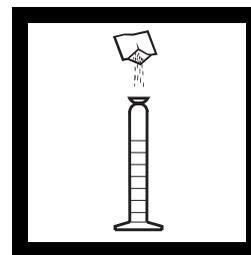
The display will show **mg/L, NO<sub>3</sub>-N** and the **ZERO** icon.

*Note:* For alternate forms (NO<sub>3</sub>), press the **CONC** key.



**3.** Fill a 25-mL graduated mixing cylinder to the 15-mL mark with sample.

*Note:* Adjust the pH of stored samples before analysis.

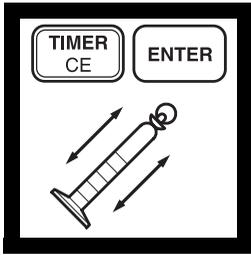


**4.** Add the contents of one NitraVer 6 Nitrate Reagent Powder Pillow to the cylinder. Stopper.

*Note:* It is necessary to remove **all** the powder from the foil pillow. Tap the pillow until no more powder pours out. Be sure to remove powder from the corners of the pillow.

\* Seawater requires a manual calibration; see Interferences.

# NITRATE, Low Range, continued

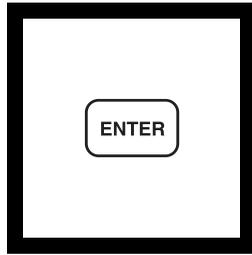


5. Press:

## TIMER ENTER

A 3-minute reaction period will begin. Shake the cylinder vigorously throughout this three minute period.

*Note: Shaking time and technique influence color development. For most accurate results, analyze a standard solution several times and adjust the shaking time to obtain the correct result.*

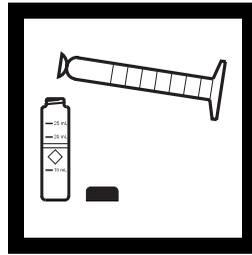


6. When the timer beeps, the display will show: **2:00 TIMER 2**

Press: **ENTER**

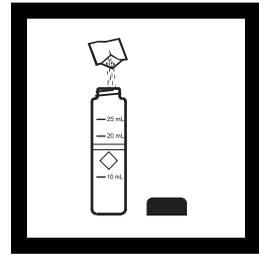
A 2-minute reaction period will begin.

*Note: A deposit will remain after the powder dissolves and will not affect results.*



7. When the timer beeps, pour 10 mL of the sample into a sample cell.

*Note: Do not transfer any cadmium particles.*



8. Add the contents of one NitriVer 3 Nitrite Reagent Powder Pillow to the sample cell (the prepared sample). Cap the cell and shake gently for 30 seconds.

*Note: A pink color will form if nitrate is present.*

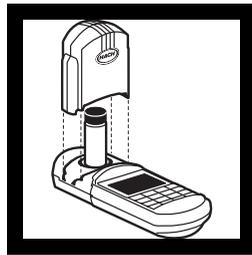


9. The display will show: **15:00 TIMER 3**

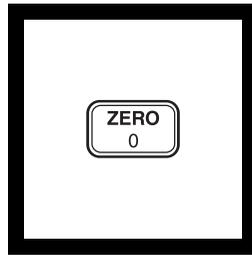
Press: **ENTER**

A 15-minute reaction period will begin.

Fill another sample cell (the blank) with 10 mL of sample.



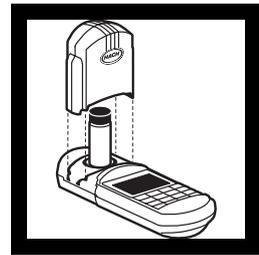
10. When the timer beeps, place the blank into the cell holder. Tightly cover the sample cell with the instrument cap.



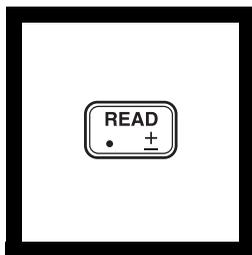
11. Press: **ZERO**  
The cursor will move to the right, then the display will show:

**0.00 mg/L NO<sub>3</sub>-N**

*Note: If Reagent Blank Correction is on, the display may flash "limit". See Section 1.*



12. Place the prepared sample into the cell holder. Tightly cover the sample cell with the instrument cap.



**13. Press: READ**

The cursor will move to the right, then the result in mg/L NO<sub>3</sub><sup>-</sup>-N (or alternate form) will be displayed.

*Note: Standard Adjust may be performed using a prepared standard (see Section 1).*

*Note: Rinse the sample cell and cylinder immediately after use to remove all cadmium particles.*

*Note: See Pollution Prevention and Waste Management for proper disposal of cadmium.*

---

### Sampling and Storage

Collect samples in clean plastic or glass bottles. Store at 4 °C (39 °F) or lower if the sample is to be analyzed within 24 to 48 hours. Warm to room temperature before running the test. For longer storage periods, adjust sample pH to 2 or less with sulfuric acid, ACS (about 2 mL per liter). Sample refrigeration is still required.

Before testing the stored sample, warm to room temperature and neutralize with 5.0 N Sodium Hydroxide Standard Solution. Do not use mercury compounds as preservatives. Correct the test result for volume additions; see *Correction for Volume Additions (Section 1)* for more information.

# NITRATE, Low Range, continued

---

## Accuracy Check

### Standard additions Method

- a) Fill three 25-mL graduated mixing cylinders with 15 mL of sample.
- b) Snap the neck off a Nitrate Nitrogen Ampule Standard Solution, 12.0 mg/L  $\text{NO}_3^-$ -N.
- c) Using the TenSette Pipet, add 0.1, 0.2, and 0.3 mL of the standard to the three samples. Stopper and mix well.
- d) Analyze each sample as described above. The nitrate nitrogen concentration should increase 0.08 mg/L for each 0.1 mL of standard added.
- e) If these increases do not occur, see *Standard Additions* (Section 1) for more information.

### Standard Solution Method

Prepare a 0.20 mg/L nitrate nitrogen standard by diluting 2.00 mL of a 10.0 mg/L Nitrate Nitrogen Standard Solution to 100.0 mL with deionized water. Use this standard in place of sample in Step 3.

### Standard Adjust

To adjust the calibration curve using the reading obtained with the 0.20-mg/L standard solution, press the **SETUP** key and scroll (using the arrow keys) to the STD setup option. Press **ENTER** to activate the standard adjust option. Then enter **0.20** to edit the standard concentration to match that of the standard used. Press **ENTER** to complete the curve adjustment. If you are using a reagent blank correction, the blank correction should be entered before the Standard Adjust feature is entered. See *Section 1, Standard Curve Adjustment* for more information.

## Method Performance

### Precision

In a single laboratory using a standard solution of 0.25 mg/L nitrate nitrogen ( $\text{NO}_3^-$ -N) and two representative lots of reagent with the instrument, a single operator obtained a standard deviation of  $\pm 0.03$  mg/L nitrate nitrogen.

### Estimated Detection Limit

The estimated detection limit for program 55 is 0.01 mg/L  $\text{NO}_3^-$ -N. For more information on the estimated detection limit, see *Section 1*.

# NITRATE, Low Range, continued

## Interferences

Interfering Substance	Interference Levels and Treatments
Calcium	100 mg/L
Chloride	Chloride concentrations above 100 mg/L will cause low results. The test may be used at high chloride concentrations (seawater) but a calibration must be done using standards spiked to the same chloride concentration.
Ferric iron	All levels
Nitrite	All levels: This method measures both the nitrate and nitrite in the sample. If nitrite is present, the nitrite nitrogen test Program 60 should be done on the sample. Pretreat the nitrate nitrogen sample with the following pretreatment. Then subtract the amount of nitrite found from the results of the LR nitrate nitrogen test using the pretreated sample. <ol style="list-style-type: none"><li>1. Add 30-g/L Bromine Water dropwise to the sample in Step 3 until a yellow color remains. Mix after each drop.</li><li>2. Add one drop of 30-g/L Phenol Solution to destroy the yellow color.</li><li>3. Proceed with the LR Nitrate procedure.</li></ol>
pH	Highly buffered samples or extreme sample pH may exceed the buffering capacity of the reagents and require sample pretreatment.
Strong oxidizing and reducing substances	Interfere at all levels

## Summary of Method

Cadmium metal reduces nitrates present in the sample to nitrite. The nitrite ion reacts in an acidic medium with sulfanilic acid to form an intermediate diazonium salt which couples to chromotropic acid to form a pink-colored product.

## Pollution Prevention and Waste Management

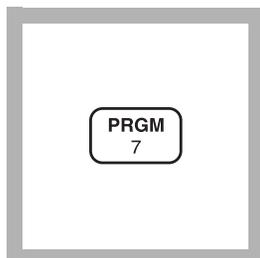
NitaVer 6 contains cadmium metal. Both samples and reagent blanks will contain cadmium (D006) at a concentration regulated as hazardous wastes by the Federal RCRA. Do not pour these solutions down the drain. See *Section 3* for more information on proper disposal of these materials.

**IRON, TOTAL (0 to 3.00 mg/L)**

For water, wastewater, and seawater

**FerroVer Method (Powder Pillows or AccuVac Ampuls)**

USEPA approved for reporting wastewater analysis (digestion is required; see Section 2\*)



1. Enter the stored program number for iron (Fe) powder pillows.

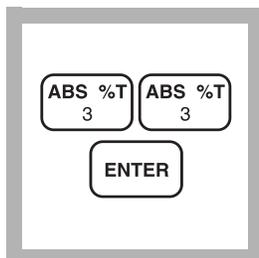
Press: **PRGM**

The display will show:

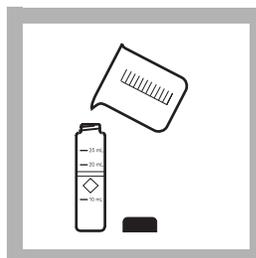
**PRGM ?**

*Note: Determination of total iron requires a digestion prior to analysis (see Section 2).*

*Note: Adjust pH of stored samples before analysis.*

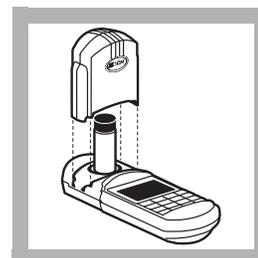


2. Press: **33 ENTER**  
The display will show **mg/L, Fe** and the **ZERO** icon.

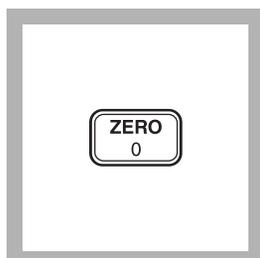


3. Fill a clean sample cell with 10 mL of sample (the blank).

*Note: For turbid samples, treat the blank with one 0.1-gram scoop of RoVer Rust Remover. Swirl to mix.*

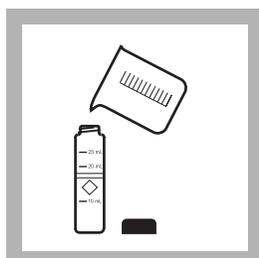


4. Place the blank into the cell holder. Tightly cover the sample cell with the instrument cap.



5. Press: **ZERO**  
The cursor will move to the right, then the display will show:

**0.00 mg/L Fe**

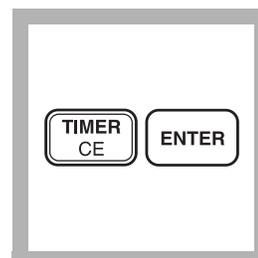


6. Fill another sample cell with 10 mL of sample.



7. Add the contents of one FerroVer Iron Reagent Powder Pillow to the sample cell (the prepared sample). Cap and invert to dissolve the reagent powder.

*Note: Accuracy is not affected by undissolved powder.*



8. Press: **TIMER ENTER**  
A three-minute reaction period will begin.

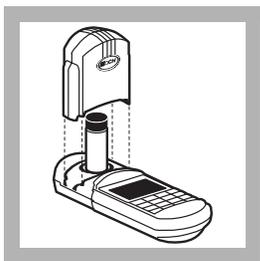
*Note: An orange color will form if iron is present.*

*Note: Samples containing visible rust should be allowed to react at least five minutes.*

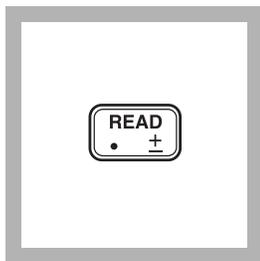
\* Federal Register, 45 (126) 43459 (June 27, 1980). See also 40 CFR, part 136.3, Table IB.

## IRON, TOTAL, continued

---



**9.** Place the prepared sample into the cell holder. Tightly cover the sample cell with the instrument cap.

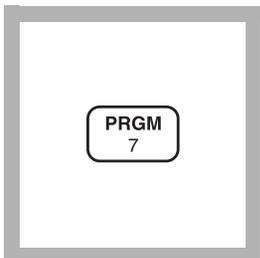


**10.** Press: **READ**

The cursor will move to the right, then the result in mg/L iron (Fe) will be displayed.

*Note: Standard Adjust may be performed using a prepared standard (see Section 1).*

### Using AccuVac Ampuls



**1.** Enter the stored program number for iron (Fe), AccuVac ampuls.

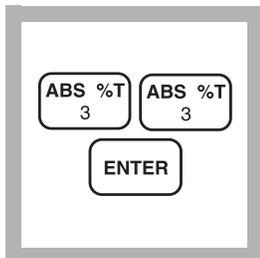
Press: **PRGM**

The display will show:

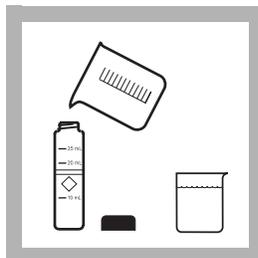
**PRGM ?**

*Note: Adjust pH of stored samples before analysis.*

*Note: Determination of total iron requires a digestion prior to analysis (see Section 2).*

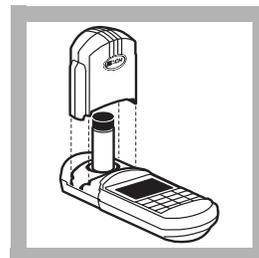


**2.** Press: **33 ENTER**  
The display will show **mg/L, Fe** and the **ZERO** icon.



**3.** Fill a sample cell (the blank) with at least 10 mL of sample. Collect at least 40 mL of sample in a 50-mL beaker.

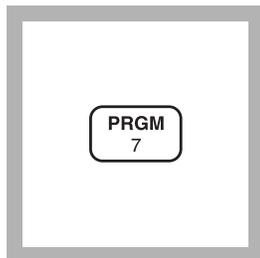
*Note: For turbid samples, treat the blank with one 0.1 g scoop of RoVer Rust Remover. Swirl to mix.*



**4.** Place the blank into the cell holder. Tightly cover the sample cell with the instrument cap.

**PHOSPHORUS, REACTIVE (0 to 2.50 mg/L PO<sub>4</sub><sup>3-</sup>)** For water, wastewater, seawater**(Also called Orthophosphate) PhosVer 3 (Ascorbic Acid) Method\***

(Powder Pillows or AccuVac Ampuls) USEPA Accepted for wastewater analysis reporting\*\*

**Using Powder Pillows**

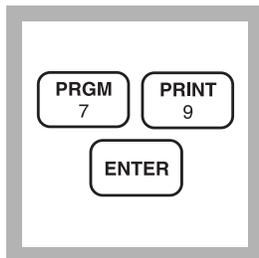
**1.** Enter the stored program number for reactive phosphorus, ascorbic acid method.

Press: **PRGM**

The display will show:

**PRGM ?**

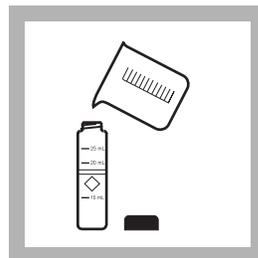
*Note: For most accurate results, perform a Reagent Blank Correction using deionized water (see Section 1).*



**2.** Press: **79 ENTER**

The display will show **mg/L, PO<sub>4</sub>** and the **ZERO** icon.

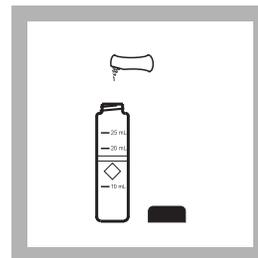
*Note: For alternate forms (P, P<sub>2</sub>O<sub>5</sub>), press the **CONC** key.*



**3.** Fill a sample cell with 10 mL of sample.

*Note: For samples with extreme pH, see Interferences following these steps.*

*Note: Clean glassware with 1:1 HCl. Rinse again with deionized water. Do not use detergents containing phosphates to clean glassware.*



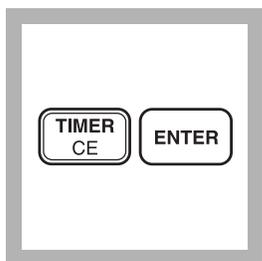
**4.** Add the contents of one PhosVer 3 Phosphate Powder Pillow for 10-mL sample to the cell (the prepared sample). Shake for 15 seconds.

*Note: A blue color will form if phosphate is present.*

\* Adapted from *Standard Methods for the Examination of Water and Wastewater*.

\*\* Procedure is equivalent to USEPA method 365.2 and Standard Method 4500-PE for wastewater.

## PHOSPHORUS, REACTIVE, continued



5. Press:

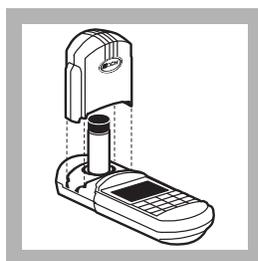
**TIMER ENTER**

A two-minute reaction period will begin. Perform Steps 6-8 during this period.

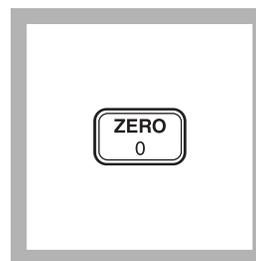
*Note: If the acid-persulfate digestion was used, an 8-10 minute reaction period is required.*



6. Fill another sample cell with 10 mL of sample (the blank).



7. Place the blank into the cell holder. Tightly cover the sample cell with the instrument cap.

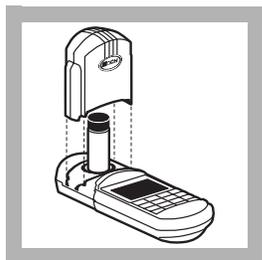


8. Press: **ZERO**

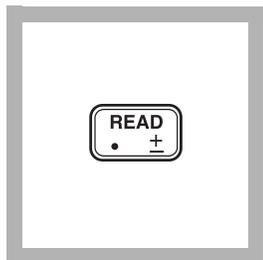
The cursor will move to the right, then the display will show:

**0.00 mg/L PO<sub>4</sub>**

*Note: If Reagent Blank Correction is on, the display may flash "limit". See Section 1.*



9. After the timer beeps, place the prepared sample into the cell holder. Tightly cover the sample cell with the instrument cap.



10. Press: **READ**

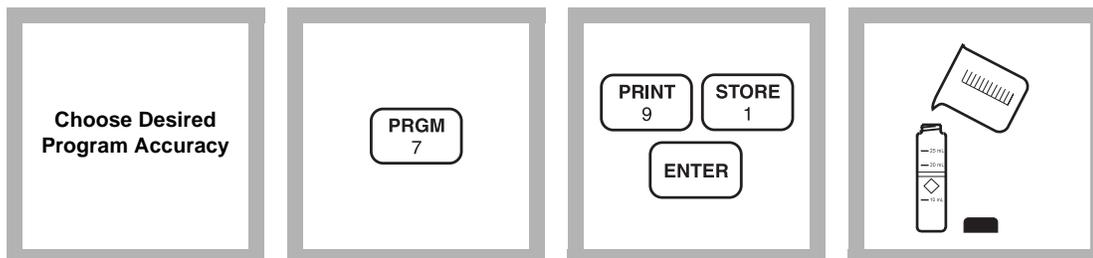
The cursor will move to the right, then the result in mg/L phosphate (PO<sub>4</sub><sup>3-</sup>) will be displayed.

*Note: Standard Adjust may be performed using a 2.0-mg/L PO<sub>4</sub><sup>3-</sup> standard; see Section 1.*

**SULFATE (0 to 70 mg/L)**

For water, wastewater, and seawater

**SulfaVer 4 Method\*** (Powder Pillows or AccuVac Ampuls); USEPA accepted for reporting wastewater analysis\*\*

**Using Powder Pillows**

**1.** A User-Entered Calibration is necessary to obtain the most accurate results. See the *User Calibration* section at the back of this procedure. Program 91 can be used for process control or applications where a high degree of accuracy is not needed.

*Note: The nature of turbidimetric tests and reagent lot variation requires user calibration for best results.*

**2.** Enter the stored program number for sulfate ( $\text{SO}_4^-$ ).  
Press: **PRGM**  
The display will show:  
**PRGM ?**

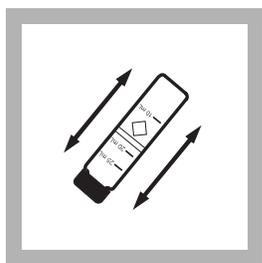
**3.** Press: **91 ENTER** or the program number selected for a user-entered calibration.  
The display will show **mg/L, SO4** and the **ZERO** icon.

**4.** Fill a clean sample cell with 10 mL of sample.  
*Note: Filter highly turbid or colored samples. Use filtered sample in this step and as the blank.*

\* Adapted from *Standard Methods for the Examination of Water and Wastewater*.

\*\* Procedure is equivalent to USEPA method 375.4 for wastewater.

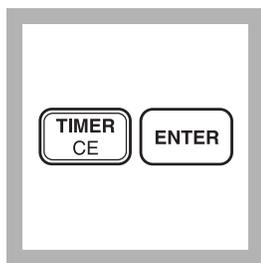
## SULFATE, continued



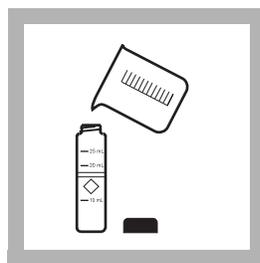
**5.** Add the contents of one SulfaVer 4 Sulfate Reagent Powder Pillow to the sample cell (the prepared sample). Cap the cell and invert several times to mix.

*Note:* A white turbidity will develop if sulfate is present in the sample.

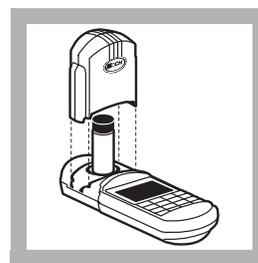
*Note:* Accuracy is not affected by undissolved powder.



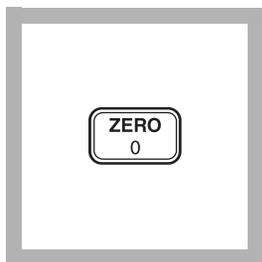
**6.** Press: **TIMER ENTER**  
A 5-minute reaction period will begin. Allow the cell to stand undisturbed.



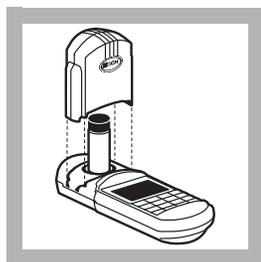
**7.** After the timer beeps, fill a second sample cell with 10 mL of sample (the blank).



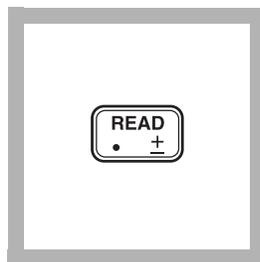
**8.** Place the blank into the cell holder. Tightly cover the sample cell with the instrument cap.



**9.** Press: **ZERO**  
The cursor will move to the right, then the display will show:  
**0 mg/L SO<sub>4</sub>**



**10.** Within five minutes after the timer beeps, place the prepared sample into the cell holder. Tightly cover the sample cell with the instrument cap.



**11.** Press: **READ**  
The cursor will move to the right, then the result in mg/L sulfate will be displayed.

*Note:* If Program 91 is used, use of the Standard Adjust is highly recommended. See Accuracy Check.

*Note:* Clean the sample cells with soap and a brush.

# CHEMICAL ANALYSIS INFORMATION, continued

---

## Using Sample Cells

### Orientation of Sample Cells

Two round sample cells are shipped with the DR/820, DR/850 and DR/890. They are marked with 10-, 20- and 25-mL fill lines which may be used to measure the sample volume unless the procedure instructs you to use other glassware to measure the sample volume.

To minimize variability of measurements using a particular cell, always place the cell into the cell holder with the same orientation. The cells are placed in the instrument with the fill marks facing the user.

In addition to proper orientation, the sides of the cells should be free of smudges, fingerprints, etc. to ensure accurate readings. Wipe the sides of the cells with a moist cloth followed by a dry soft cloth to clean the surface before taking measurements.

### Care of Hach Sample Cells

Store sample cells in their boxes when not in use to protect them from scratching and breaking. It is good laboratory practice to empty and clean sample cells after analyses are complete--avoid leaving colored solutions in the cells for extended periods of time. Finish the cleaning procedure with a few rinses of deionized water and allow to dry. Individual procedures often recommend specific cleaning methods.

### Cleaning Sample Cells

Most laboratory detergents can be used at recommended concentrations. Neutral detergents such as Neutracon are safer if regular cleaning is required, as in the case of protein residues.

If using a detergent, you can speed cleaning by increasing the temperature or using an ultrasonic bath.

Rinsing is more efficient when using deionized water.